Nursing Guidelines on the Care of Neonates and Infants with Thermoregulation Instability

2nd Edition

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1.0 Thermoregulation Overview

Introduction
Thermoregulation is a critical function which is closely linked to the survival of preterm infants, sick neonates and infants. The role of the nurse is crucial in limiting heat loss and providing the correct thermal environment. The nurse has a responsibility to ensure that heat loss is minimised and that thermal conditions are stable for the infant. Monitoring accurate core and skin temperatures are vital. It is fundamental to understand the mechanisms of heat production and heat loss pertaining to the preterm infant. The nurse must also have a thorough knowledge of the equipment available and how to use it safely and appropriately (Fellows 2010).

Definition of Thermoregulation
Neutral thermal environment has been defined as when the infants’ temperature does not change, with a stable metabolic state where minimal rates of oxygen consumption or energy expenditure occur (Roncoli and Medoff-Cooper 1992, Knobel and Holditch-Davis 2007, Brown and Landers 2011). Sauer et al (1984) recommend that neutral thermal environment is best achieved when infants can maintain a core temperature at rest of between 36.7 and 37.3 °C allowing for a change of only 0.2-0.3 °C per hour.

World Health Organisation (WHO) Classification (Thermal Control of the Newborn)
- 36-36.4 °C = Cold stress. = Mild Hypotherma, cause for concern.
- < 36.0 °C = Moderate hypothermia. Dangerous requires immediate warming of the infant.

Indications for Assisted Thermoregulation
Normal body temperature symbolises the optimum thermal condition required to maintain internal body function. Thermoregulation provides a balance between heat generation and heat loss therefore maintaining normal body temperature. Thermoregulation is a vital body function, which is reflective of physiological maturity. The neonate (0-28 days of life) has to instigate thermoregulation at birth. However, if born prematurely the physiological pathways have not yet fully developed to initiate this action. Furthermore sufficient reserves are not present in the premature infant to maintain thermal stability without compromising other body systems (Fellows, 2010). Neonates are more prone to poor thermoregulation due to:
- Intrauterine temperature has been constant unlike the external environment
- High metabolic rate
- Large surface to body mass ratio
- Large head (25% heat loss)
- Reduced/poor insulation with lack of subcutaneous fat
- Permeable skin
- Immature hypothalamus, central nervous system, vasomotor control
- Neonates have limited shivering response and may lack the ability to produce heat. Whilst the preterm infant has no shivering response at all.
- Reduced energy stores
- Dependent /communication difficulties
- Poor muscle tone and inability to position own body

Infants Most at Risk
- Low Gestational age (LGA)
- Extremely low birth weight (ELBW) infants
- Cardio-respiratory, neurological & endocrine disease
- Congenital abnormalities, e.g. gastroschisis, exomphalos
- Sedation, muscle relaxation
- Hypoglycaemia
- Caesarean (LSCS) delivery (in the immediate postnatal period)

Complications Associated with Thermo Instability
Heat production takes place through oxidation of metabolic substrates (metabolism), non-shivering thermogenesis, muscle activity and vasoconstriction.

Excessive Heat Gain can lead to:
- Increased fluid loss
- Hypernatraemia
- Increased jaundice
- Recurrent apnoea
- Increased neonatal mortality
Heat Loss takes place through conduction, convection, radiation and evaporation.

- **Conduction** accounts for 10-15% of heat loss and occurs when objects are placed in contact with each other, e.g. a warm infant is placed on a cool surface. Heat is then transferred from one to the other i.e. cold x-ray plate, stethoscope or weighing scales.

- **Convection** involves heat loss due to the movement of air at the skin surface i.e. draughts. Cool environmental temperature. open incubator ports. It is the second most common mode of heat loss in the preterm and term infant.

- **Radiation** is the transfer of heat energy from the exposed surface of the infant to the surrounding surfaces i.e. cold incubator walls. In the term infant it accounts for 60% of heat loss.

- **Evaporation** is the insensible water loss from the skin surface and the respiratory mucosa. It accounts for 60% of heat loss in the preterm infant. (Roncoli and Medoff-Cooper 1992, GOSH 2008, EOENBG 2011).

**Excessive Heat Loss** contributes in particular to:

- **Acidosis** due to metabolism of fatty acids and increased lactic acid accumulation.

- **Increased oxygen consumption** causing hypoxemia / hypoxia, desaturation due to anaerobic metabolism from pulmonary hypertension and also vasoconstriction and decreased delivery of oxygen to tissues.

- **Hypoglycaemia** due to increased metabolic rate, glucose utilisation and depletion of glycogen stores (Karlsen 2006, Knobel and Holditch-Davis 2007).

REMEMBER allowing the infant to get cold and stay cold, even mild hypothermia / cold stress increases neonatal mortality and morbidity (Fellows 2010).

“Preventing hypothermia is much easier than overcoming the detrimental effects of hypothermia once they have occurred” (Karlsen 2006:58).

**Hypothermia can also Lead to:**

- Impaired surfactant synthesis and efficiency
- Tachyypnoea resulting from an increased need for oxygen due to increased metabolism
- Bradycardia due to hypoxia
- Pale mottled skin due to poor perfusion and hypoxia
- Cold extremities with poor perfusion from peripheral vasoconstriction which occurs as the infant tries to conserve heat
- Poor feeding, with gastric distension or large aspirates / vomiting
- Failure to gain weight or weight loss due to increased metabolism, non-shivering heat production and increase in consumption of calories
- Altered blood coagulopathy and risk of bleeding and thrombocytopenia
- Increased risk of infection
- Neurological damage due to hypoglycaemia
- Restlessness, irritability, hypotonia and lethargy
- Altered cerebral blood flow which may predispose to intraventricular haemorrhage
- Huperbilrubinaemia and kernicterus
- Acute renal failure as glomerular filtration rate (GFR) declines due to poor perfusion

**Equipment Required for Thermoregulation**

- Incubator / Radiant Warmers / Giraffe Omnibed
- Humidity
- Temperature monitoring device (Skin and Core)
- Clean linen

**Scope of the Guidelines**

Applies to all infants up to and < one year of life including preterm’s, neonates and all low birth weight (LBW) infants <2.5kgs. For the purpose of this guideline a preterm is defined as an infant < 37/40 weeks gestation

A neonate or term infant is an infant of 37/40 gestation up to and including 28 days of life.

**Nursing Documentation**

The nurse is responsible for accurate recording and documentation of all nursing care/ observations to maintain accountability and continuity of patient care (An Bord Altranais 2002).
### 2.0 Thermoregulation within Incubators

The neonatology team in OLCHC recommend that all infants < 2kgs are nursed in an incubator in PICU / ward areas.

**Specific Equipment:** Incubator +/- humidity depending on gestational age. Hat. Mittens, booties as indicated.

**NB:** Never turn off an incubator with the infant still inside. Always reduce incubator temperature. Opening portholes to cool incubator is ineffective as incubator only works harder to produce set heat.

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<thead>
<tr>
<th>ACTION</th>
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<tr>
<td>Incubator</td>
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<tr>
<td>All preterm infants, low birth weight, or sick term neonates transferred to PICU are admitted into a pre-warmed incubator.</td>
<td>Sick term infants and premature infants have difficulty in self regulation of temperature. Incubators help reduce heat loss by conduction, radiation and secure heat gain. Incubators also help prevent cross infection, promote minimal handling, aid with noise reduction and enable close observation of the sick neonate (Fellows 2010).</td>
</tr>
<tr>
<td>It is preferable to keep preterm infants and low birth weight term/sick neonates in incubators whilst in the PICU environment and if &lt;2kgs within the ward areas.</td>
<td>To prevent cross infection, cold stress and other effects of poor thermoregulation in the newborn sick infant and premature infants (O’Connor 2010)</td>
</tr>
<tr>
<td>Provide explanation and give continued ongoing support to parents / guardians. Promote maternal and parental bonding.</td>
<td>To increase parental confidence, autonomy and allow bonding to take place (Fellows 2010, Trigg and Mohammed 2010).</td>
</tr>
<tr>
<td>Prepare the incubator and preheat, in preparation for the infant</td>
<td>To prepare for the infants admission (Trigg and Mohammed 2010).</td>
</tr>
<tr>
<td>Close all windows and doors and ensure privacy.</td>
<td>To ensure the infant is placed in a warm environment and to prevent draughts and cold stress. Also to prevent heat loss due to convection (Dougherty and Lister 2008, Trigg and Mohammed 2010).</td>
</tr>
<tr>
<td>Position incubator out of direct sunlight and away from radiator and ensure wheels are locked in position.</td>
<td>To ensure the incubator / infant isn’t subjected to temperature flux from the environment and to ensure health and safety issues are incorporated (Trigg and Mohammed 2010).</td>
</tr>
<tr>
<td>Ensure the incubator is safely situated without obstruction from furniture / equipment and away from the walls so that both side doors can be freely let down to allow access to the infant by staff members.</td>
<td>To allow access to the infant from both sides of the incubator in case of an emergency i.e. resuscitation.</td>
</tr>
<tr>
<td>The air temperature mode should be used to set the incubator pre-warmed to:</td>
<td>When the incubator temperature is greater than the infants temperature this will secure heat gain and help to reduce heat loss by conduction and radiation. It will also ensure the infant nursed in incubator isn’t subjected to temperature fluctuations from the environment and to ensure health and safety issues are incorporated. The term infant has a lower temperature set to avoid overheating the infant (St Mary’s Hospital 2008a, Trigg and Mohammed 2010, EOENBG 2011).</td>
</tr>
<tr>
<td>• 37 °C. - Preterm infant</td>
<td>To prevent cold stress and provide comfort for the infant (EOENBG 2011).</td>
</tr>
<tr>
<td>• 35 °C. - Term infant</td>
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All temperature probes should be pre warmed in the incubator prior to attaching to infant.
Once the infant is placed in the incubator, the air temperature should then be reduced and set accordingly to maintain infant’s temperature within a neutral thermal temperature, i.e. 36.7-37.3°C. (Appendix 3).

Monitor core and peripheral temperatures continuously and document same i.e.:
- PICU’s. in the preterm or LBW infant < 1.8kgs
- Intubated, unstable, inotrope dependent
- Until the infant no longer needs to be in an incubator and has successfully transferred to a cot. Infant is considered clinically stable with expected weight gain and on full feeds.

In exceptionally rare cases within ward areas, some sick neonate may require skin and core temperature and ECG monitoring, as per medical team.

As infant matures and becomes more stable 4 hourly clinical observation, assessment and documentation of core and peripheral temperature is acceptable within the PICU and ward areas. Core temperatures can be monitored age appropriately i.e. tempadot. Peripheral temperatures can now be monitored via touch / feel i.e., “warm to toes” and, “warm to finger tips” method. The infant should have more frequent monitoring/ recording of core / peripheral temperatures if their condition becomes unstable / deteriorates as clinically indicated.

Close monitoring of central / peripheral temperature and incubator temperature is necessary when undertaking care to preterm infant and to interpret if the neutral thermal environment is compromised.

**Core temperature** (while in PICU) should be monitored using:
- Skin temperature probe between mattress – skin (intrascapular) attached to a cardiac monitor
- Rectal temperatures should be avoided where possible, however they may be indicated PICU (ONLY), i.e.
  - Post cardiac surgery, meningococcal septicaemia, sepsis, cooling of infant to protect brain i.e. asphyxia.

NB: Where used rectal temperature monitoring should be of short duration (<24hrs) unless best practice indicates otherwise.

**Peripheral temperature** monitoring (while in PICU):
- Peripheral skin probe attached to sole of foot
- Used routinely for all premature infants and sick neonates.

Infants nursed in “air control mode” have a more stable thermo-regulated environment and less variance between core and peripheral temperatures (Ducker et al 1985, Mok et al 1991, Boyd and Lenhart 1996).

To maintain infant in a neutral thermal environment.

To ensure early detection and timely intervention for temperature fluctuations (Fellows 2010). A temperature gradient >2°C between skin (peripheral) and core may be an early indication of cold stress as the infant tries to minimise heat loss and should be investigated. Core temperatures which are measured from abdomen, or axilla whilst mainly accurate, may be subject to heat fluctuations from surrounding environment (Lyon 2004, Sherman et al 2006, Fellows 2010).

Incubator temperatures fall during care when portholes or incubator doors are open with subsequent drop in the preterm infant’s / sick neonates central and peripheral temperature subjecting them to the risk of cold stress (Mok et al 1991).

Probe between scapular and non-conducting mattress is very accurate (Lyon 2004, EOENBG 2011).

Rectal temperatures are extremely invasive and may be unreliable. A rectal temperature probe predisposes to rectal polyps (Fellows 2010).

In the 1st 2-3 days of life the pre-term infant is poikilothermic i.e. adopts the temperature of the environment. He then develops the ability to peripherally vasoconstrict, shunting blood to the core when challenged thermally (Lyon et al 1997). Peripheral temperature is also an early indication of cold stress and also poor perfusion (EOENBG 2011).
| Preterm infant’s the core temperature should be maintained between 36.7- 37.3 °C |
| Monitor and document incubator temperature hourly (PICU).  2-4 hourly if the infant is clinically stable at ward level. |
| Hat, mittens and booties should be used for infants. |
| Within the PICU’s the temperature alarm limits on cardiac monitor should be tightly set, i.e. 0.2 °C above and below accepted parameters. |
| Oxygen/air gases should always be humidified and warmed. |
| The ventilator temperature probe sits inside the incubator and must be shielded from environmental flux by the use of heat reflective shield. |
| When infant has stabilised, dressing the infant fully whilst still in the incubator is encouraged as clinically indicated. |
| Temperature monitoring should be continuous: |
| • PICU’s |
| • First few days after transfer to a cot |
| The initial set incubator temperature is reduced in preparation for transfer to a cot. |
| **Safety** |
| Care should be taken when placing covers over incubators. |
| Items or electrical equipment should not be placed on the top or in the incubator |
| Incubators should be changed weekly and sent for cleaning and routine maintenance checks. More frequently if soiled or if the infant is septic, as clinically indicated |
| To help maintain temperature of the infant and reduce heat loss (Sauer et al 1984, British Columbia 2003, EOENBG 2011). |
| To observe the frequency of changes to the incubator temperature which may indicate that extra energy is being expended by the infant (GOSH 2008, EOENBG 2011). |
| The infants head has a large surface area in proportion to size and is vulnerable to heat loss. (Lyons and Oxley 2001, Knobel et al 2009). |
| To assist in the early detection of temperature variations and potential complications of same (Blissinger and Annibale 2010, Trigg and Mohammed 2010). |
| This decreases insensible water and heat loss from respiratory tract. Endotrachial tubes bypass the natural humidification and filtering systems. |
| If the extension tubing is used the gases may cool before reaching the infant as the heating wire only goes as far as the temperature probe. |
| The sick and/premature infant is less at risk of cold stress once stabilised but is still at risk (Fellows, 2010). |
| To ensure early detection and timely intervention for temperature fluctuations. |
| The infant becomes more mature; condition improves, is maintaining own temperature and ready to be dressed in preparation for transfer to cot. |
| Whilst this may minimise light to the infant it may also reduce visibility and mimic the day / night effect (Fielder and Moseley 2000, Lee et al. 2005). The nurse must be able to assess the infant’s condition at all times. |
| Placing the item (i.e. feeding bottles) on top of the incubator can be very noisy and cause undue stress for the infant (Reid and Freer 2003). |
| NB: Noise created outside the incubator is amplified greatly inside the incubator. |
| To minimise the risk of infection (OLCHC 2006, 2008a) |
3.0 Thermoregulation in Giraffe Omnibed (PICU Only)

Giraffe Omnibed has two modes:
- Radiant warmer
- Incubator

Specific Equipment Required: Giraffe Omnibed

Patient indicators
Preterm infants <1500 grams and < 32 weeks gestation.

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<td>All preterm infants &lt;1500 grams should be admitted to a prewarmed Giraffe OmniBed (where possible).</td>
<td>The Giraffe OmniBed is a radiant warmer that can be converted to an incubator, once the infant has been stabilised. This eliminates the stress of moving critically-ill infants from bed to bed. The mattress in the giraffe rotates facilitating proper positioning of the infant for all types of procedure without having to physically move the infant (GE Healthcare 2010).</td>
</tr>
<tr>
<td>Keep parents / guardians informed and explain all equipment as clinically indicated.</td>
<td>To reduce parental stress and provide support though the use of information (Trigg and Mohammed 2010).</td>
</tr>
<tr>
<td>On admission, infants should be nursed under the radiant warmer in servo mode.</td>
<td>This mode allows stabilisation of the infant’s temperature and facilitates examination and procedural interventions.</td>
</tr>
<tr>
<td>Servo mode should be utilised when used as radiant warmer instead of an incubator.</td>
<td>To maintain the infants’ temperature constant and prevent cold stress (Lyon 2004).</td>
</tr>
<tr>
<td>Keep the sides of the warmer up where possible</td>
<td>To prevent the infant from falling from the Giraffe and maintain a warm environment by preventing unnecessary draughts (Trigg and Mohammed 2010)</td>
</tr>
<tr>
<td>The Giraffe OmniBed should be converted from radiant warmer to incubator mode once the infants’ temperature has been stable for an hour and nursing cares / procedural interventions have been completed.</td>
<td>Incubators provide a more stable temperature (thermal neutral environment) in comparison to radiant warmers and reduces the incidence of trans-epidermal water loss (TEWL). In newborn very extremely low birth weight (ELBW) preterm infants, evaporation of water from the skin is an important mode of fluid loss (Agren et al 1998, Lyon 2004).</td>
</tr>
<tr>
<td>Servo Mode</td>
<td>This is to allow maintenance of a thermal neutral environment and easy conversion between radiant warmer and incubator in emergency situations without having to adjust settings thus preventing the infant from developing cold stress.</td>
</tr>
<tr>
<td>Should be used for all critically ill infants who are nursed naked (nappy only) in an incubator.</td>
<td>Humidified warm air reduces TEWL and may decrease the infant’s air temperature requirements. TEWL are high in the immature baby. The 26 week infant, on day 1 of life, can lose over 50 kilocalories / kg via evaporation compared with less than 5 kilocalories / kg in the term infant (Lyon 2004).</td>
</tr>
<tr>
<td>Once changed to incubator mode, humidity should be commenced as clinically indicated (Section 5).</td>
<td>Air mode can be adjusted by the nurse to maintain a stable body temperature in infants in accordance with the amount of clothing applied.</td>
</tr>
<tr>
<td>Air Mode</td>
<td>This creates a strong blanket of air to serve as a barrier against cool air entry when a porthole or door is opened. In accordance with recommended positioning for infants</td>
</tr>
<tr>
<td>Should only be used for the stable dressed infant nursed in an incubator as clinically indicated.</td>
<td>During nursing care or procedural interventions the ‘Boost Air Curtain’ should be used (Appendix I). The infant should be placed directly on the linen covered</td>
</tr>
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</table>
mattress and surrounded by boundaries (nesting).

All temperature probes should be checked at the start of every shift and an axilla temperature taken to check correlation of core temperature probe. 
- If the infant's core temperature changes > 0.5 °C an axilla temperature should be taken and the position of the servo probe checked.

The Servo temperature probe should be positioned either on the infants:
- Middle of abdomen, if nursed supine
- The flank of the back, if nursed prone.

Monitor core and peripheral temperatures continuously and document same i.e.:
- PICU’s, in the preterm or LBW infant < 1.8kgs
- Intubated, unstable, inotrope dependent
- Until the infant no longer needs to be in an incubator and has successfully transferred to a cot. Infant is considered clinically stable with expected weight gain.

Close monitoring of central / peripheral temperature and incubator temperature is necessary when undertaking care to preterm infant and to interrupt if the neutral thermal environment is compromised.

Monitor and document incubator temperature hourly.

Weigh infants using the in-bed scales when nursed in the Giraffe Omnibed.

Phototherapy for infants nursed in the Giraffe Omnibed

**Hood Cover**

Used with preterm infants

NB: The hood can be raised quickly should the infant’s condition deteriorate / become unstable as clinically indicated.

Open Giraffe hood at the start of every shift as clinically indicated.

Do not use the Giraffe hood initially during an acute admission until stable.

Frequency of use should be reduced as the pre-term infant (Sweeney and Gutierrez 2002, Nair et al. 2003).

The mattress in the Giraffe is a layered Pressure-Diffusing Mattress designed to help relieve pressure points and preserve skin integrity in the critically ill infant (GE Healthcare 2007a, 2007b).

Temperature probes can become detached from the infant secondary to environmental humidity in the incubator, skin moisture, nursing care or movement of the infant. Correlation of core temperature probe to axilla temperature is to ensure that the infant is not overheating due to incorrect temperature probe placement.

If the probe is positioned on the abdomen when the infant is prone, a falsely high temperature may be recorded (Hockenberry and Wilson 2008).

To ensure early detection and timely intervention for temperature fluctuations (Fellows 2010). A temperature gradient >2 °C between skin (peripheral) and core may be an early indication of cold stress as the infant tries to minimise heat loss and should be investigated. Core temperatures which are measured from abdomen, or axilla whilst mainly accurate, may be subject to heat fluctuations from surrounding environment (Lyon 2004, Sherman et al 2006, Fellows 2010).

It has been reported that incubator temperatures fall during care when portholes or incubator doors are open with subsequent drop in the preterm infants / sick neonates central and peripheral temperature subjecting them to the risk of cold stress (Mok et al 1991).

To observe the frequency of changes to the incubator temperature which may indicate that extra energy is being expended by the infant (British Columbia 2003, GOSH 2008, EOENBG 2011).

This allows for weights to be taken and trended without removing the infant thus promoting minimal handling and preventing additional stress to the infant whilst allowing the nurse to evaluate weight gain/loss (GE Healthcare 2007c).

Phototherapy can be administered using a biliblanket, overhead lights or the Spot PT Lite (Appendix 1).

A hood cover may be used to minimise light stimulation for pre-term infants which increases infant’s stability, respiratory instability, reduces heart rate, blood pressure, respiration rate, and motor activity (Fielder and Moseley 2000).

To ensure timely access to the infant in an emergency.

To allow a thorough examination of the infant.

Hood covers can impede the close observation and visual
approaches term.

Use at night and quiet time only once infants reaches term.

Giraffe Omnibed should be changed weekly or more frequently if soiled or if the infant is septic.

monitoring of infants. The colour of the fabric can reflect off infants making it difficult to recognise a change in colour (Lee et al. 2005).

To accustom them to light (Fielder and Mosely 2000).

To allow infants distinguish between night and day (Fielder and Mosely 2000).

To minimise infection risk (OLCHC 2006, 2008a).

### 4.0 Thermoregulation in a Radiant Warmer (PICU only)

**Specific Equipment Required:** Radiant Warmer

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<tr>
<td>Nursing staff should familiarise themselves with the radiant heaters used within OLCHC by consulting the operator manual and taking direction from the technical support team within the PICUs. Bleep: 8465</td>
<td>To reduce anxiety and stress caused by hospitalisation (Trigg and Mohammed 2010).</td>
</tr>
<tr>
<td>Explanation to parents/guardians as clinically indicated.</td>
<td>To ensure the infant is placed in a warm environment and to prevent cold stress.</td>
</tr>
<tr>
<td><strong>Prepare the Environment</strong> Prepare the radiant warmer, in preparation for the infant on manual mode.</td>
<td>To keep the environment warm, draught free and also to prevent heat loss due to convection (Trigg and Mohammed 2010).</td>
</tr>
<tr>
<td>Close all windows and doors.</td>
<td>To allow access to the infant in case of an emergency.</td>
</tr>
<tr>
<td>Ensure the radiant warmer is safely situated without obstruction from furniture/equipment etc and that both side doors are free to open completely and allow access by staff members.</td>
<td>To ensure the incubator/infant isn’t subjected to temperature flux from the environment and to ensure health and safety issues are incorporated (Trigg and Mohammed 2010).</td>
</tr>
<tr>
<td>Position Radiant warmer out of direct sunlight and away from radiator and ensure wheels are locked in position.</td>
<td>Radiant warmers provide easier access to the critically ill infant. When procedures/investigations/surgery are required it can be prudent to nurse the infant in a radiant warmer. This, however, should be short term and the baby should be placed in a closed incubator as soon as possible. The Giraffe Incubator with radiant warmer option should be used in the preterm especially &lt; 30 weeks gestation. Radiant heaters can subject neonates to increased trans epidermal water loss (TEWL) and possible electrolyte imbalance, variances in thermal stability, increased oxygen consumption and handling. Also oxygen consumption increases by 8.8% under radiant warmers (LeBlanc 1982, Mok et al 1991, Davenport 1992, Sequin and Vieth 1996, Birmingham Children’s Hospital 2003)</td>
</tr>
<tr>
<td>When infants are admitted to PICU it is always preferable to nurse the infant in an incubator. However this may create challenges i.e.</td>
<td>This reduces heat loss through conduction and radiation (Ohmeda Medical 1994).</td>
</tr>
<tr>
<td>• Ventilation: High Frequency Oscillation Ventilation (HFOV)</td>
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<tr>
<td>• Gastrostisis silo bag (unrepaired gastrostisis) (Appendix 1).</td>
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<tr>
<td>Admit the infant into a pre-warmed radiant warmer using the manual mode heated to 25% power.</td>
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The manual mode, alarms every 12 minutes to alert the nurse to check the infant.

All preterm infants and sick neonates nursed in PICU should be nursed in the servo mode.

When the infant has been transferred to the radiant warmer the infants should be nursed on:
- Air mattress
- Radiant warmer mattress
- Infants less than 30wks gestation gamgee may be used if thought appropriate on an individualised basis. Use of gamgee should be discontinued at 30 weeks unless otherwise indicated by consultant or neurodevelopment physiotherapist / individualised clinical indication.

The preterm infant < 31 weeks gestation will require humidity. Therefore it is a priority that the infant is moved to a Giraffe incubator preferably or to a closed incubator as soon as possible.

In the preterm infant cling film may be placed over the bottom end of the radiant warmer as clinically indicated.

Ensure that the bedside panels are locked in position when the infant is in the warmer.

Radiant warmers increase the infants' insensible water losses especially in the low birth weight infant compared to incubators. This water loss needs to be taken into account when daily fluid requirements are calculated i.e. increased by 10 -20 % as discussed with neonatologist/ medical team.

Urinary output should be monitored closely.

**Servo Temperature Probe**

*NB: Please note that the temperature probe alarm is only active in servo mode*

In servo mode the servo skin temperature probe should be in situ.

Place the servo skin temperature probe midway centrally above the umbilicus in the direct path of radiant heat with the metal side in contact with the skin and the heat reflective foil patch facing up.  

*NB: Do not cover servo skin probe with bedding.*

Place the skin probe on infant’s back, with foil facing uppermost when the infant is nursed prone

Change servo probe site a minimum of once every shift. Care must be taken when removing or resiting adhesive pads.

Temperature probe-skin contact should be checked every 30 minutes to hour.

To prevent overheating and evaluation of preheating of the radiant warmer thereby ensuring the safety of the patient.

To maintain the infants temperature in a neutral thermal environment and prevent cold stress (Lyon 2004).

A firm mattress is needed to facilitate development. (Reid and Freer 2003).

Evaporation and water losses are higher under radiant heaters compared to incubators. The use of humidity can help reduce Transthermal epithelial water loss (TEWL) and maintain the infant’s body temperature (Kjartansson et al 1995, Flenady and Woodgate 2009, GE Healthcare 2010).

To regulate temperature gradients (hotter and cooler areas) and prevent draughts. This is due to radiant heat being applied from above the infant, the cooling effects of the mattress, TEWL, unequal internal heat generation within infant and environmental variables i.e. room temperature, room air movement, sunlight, etc (Ohmeda Medical 1994).

To prevent the infant from falling from the radiant warmer and maintain a warm environment by preventing unnecessary draughts (Trigg and Mohammed 2010).

Radiant warmers may also increase the infant's insensible water losses (Kjartansson et al 1995, Flenady and Woodgate, 2009, Fellows 2010).

This will determine accurate fluid and electrolyte balance (Fellows 2010).

(Ohmeda Medical 1994).

Brown adipose tissue (BAT) deposits are located around the neck, between the scapulae, across the clavicle line and down the sternum. It also pads the kidneys and the thoracic vessels. BAT deposits can absorb heat giving inaccurate temperature readings. The skin is very thin over bony areas and bone is not a good conductor of heat.

To prevent skin damage to delicate and immature skin (Fellows 2010).

To ensure satisfactory skin contact and early detection of servo probe lifting resulting in over heating of the infant. A 2-3 °C gap between skin and core can give an early
The servo temperature should not be relied on. Peripheral and core temperatures must always be checked separately and continuously.

The infant should be placed in a warmed incubator as soon as possible.

The radiant warmer should be changed weekly as clinically indicated.

indication of cold stress, hypovolaemia, shock, infection. In cold stress the peripheral temperature will fall before the central (Lyon 2004).

Incubators assist in preventing cross infection, promote minimal handling, aid with noise reduction and enable close observation of the sick neonate (Fellows 2010).

To minimise infection risk (OLCHC 2006, 2008a).

### 5.0 Thermoregulation and Humidity

**Specific Equipment Required:** Sterile Humidity container and sterile water

**Scope of Guideline**
- <34 week gestation infant depending on prematurity
- Infants with some skin condition i.e. scalded skin. Where humidity is incorporated for the treatment of skin conditions the level and length will be determined by neonatology/dermatology consultant.

<table>
<thead>
<tr>
<th>ACTION</th>
<th>RATIONALE &amp; REFERENCE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Offer explanation to parents / guardians</td>
<td>Providing parents/guardians with appropriate information reduces anxiety and stress caused by hospitalisation (Trigg and Mohammed 2010)</td>
</tr>
<tr>
<td>All Infants’ &lt; 31 weeks gestation should routinely be nursed in a warmed and humidified environment</td>
<td>Premature infants, due to their large surface area in relation to body weight are poorly developed and have thin skin due to lack of keratin. They may lose large amounts of heat by evaporation, which is known as trans-epidermal water loss (TEWL). The preterm infant has only a thickness of 2-3 cell layers of stratum corneum (outer most area of epidermis) especially over the abdomen, compared to the term infant who has 10-20 layers. TEWL can be up to 18 times higher in the 25/40 infant than in full term infant (Sedin <em>et al</em> 1983, Royal Prince Alfred Hospital 2009, Blissinger R.L and Annibale D.J. 2010). Each 1ml that evaporates from skin is equal to 560 kcal heat loss. TEWL is dependent on the epidermal barrier, the temperature, air speed and humidity. The use of humidity will reduce TEWL and when 100% humidity is added TEWL is stopped (Marshall 1997, Royal Prince Alfred Hospital 2009, Fellows 2010). (Queen Mothers Hospital, St. Mary’s Hospital. 2008, Heuchen and Williams 2009, Coombe Hospital 2010, O’Connor 2010)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Gestation of Delivery</th>
<th>Humidity Percentage</th>
<th>When to Discontinue</th>
</tr>
</thead>
<tbody>
<tr>
<td>28-31 weeks</td>
<td>85% humidity decreasing gradually from day 4</td>
<td>Discontinue 2 weeks after starting humidity</td>
</tr>
<tr>
<td>&lt;28 weeks</td>
<td>85% humidity decreasing gradually from day 4</td>
<td>Discontinue 3-6 weeks after starting humidity, depending on prematurity</td>
</tr>
</tbody>
</table>

Table 1: Percentage of Humidity Treatment for Preterm Infants Nursed in Incubators.

The above criteria are dependent on the clinical condition of the infant and must be in consultation with the neonatal consultant.

Use incubators with integral humidity.

The 26 week gestation infant has developed a very thin keratinised stratum corneum. Birth accelerates this maturation of the skin and the process can take between 4-6 weeks, depending on prematurity. The stratum corneum doesn’t become functionally mature until the infant is 32 – 34 weeks gestation. Humidity is considered ineffective after this time (Kalia *et al* 1998, Nonato and Lund 2001, Blissinger and Annibale 2010).
Humidity unit is sterile and Sterile Water for Irrigation (1000 mls) is used in the humidity unit which is then removed for cleaning and re-sterilisation every 24hrs. Replace with a pre-sterilised humidity unit.

Although the bedding can feel wet (damp to touch), it should only require changing once -twice a day unless soiled.

Humidity will create a thin mist on the inside of the incubator. This is acceptable and should disperse down the sides of the incubators. However if heavy rainout / excessive rainout occurs causing droplet formation which drops on the infant the humidification system / incubator should be changed and checked by the clinical engineers, Bleep 8465.

Infants should not be dressed when using humidity.

Reduce and discontinue humidity as the infant matures taking cognisance of the degree of prematurity.

Within OLCHC we have devised our humidity table (Appendix II) to incorporate humidity up to and including 34 weeks gestation

To reduce / prevent risk of colonisation by pseudomonas and other bacteria. Infection Control Team (National Maternity Hospital 2008, OLCHC 2008a)

There can be a lot of misting from high humidity levels causing bedding to feel wet but once the temperature is correct the infant will not be cold (Coombe Hospital 2009).

There is no need to wipe the inside of the incubator. Wiping can cause droplets to fall on infant causing distress. Visibility shouldn’t be a problem
The infant should be switched to another incubator in the meantime.

As clothes create a barrier to moisture created by humidity.

The literature varies but between the ages of 30-34 weeks gestation the infant’s skin is considered comparable to that of an adult and that humidity is ineffective after this stage (Kalia et al 1998, Nonato and Lund 2001, Blissinger and Annibale 2010, Fellows 2010).

To aid the weaning of humidity as the stratum corneum doesn’t become functionally mature until the infant is 32 – 34 weeks gestation in the ELBW infant (Nonato and Lund 2001, Blissinger and Annibale 2010, O’Connor, 2010).

6.0 Thermoregulation Control when Transferring an Infant from an Incubator to an Open Cot

<table>
<thead>
<tr>
<th>ACTION</th>
<th>RATIONALE &amp; REFERENCE</th>
</tr>
</thead>
<tbody>
<tr>
<td>The transfer should be a gradual process.</td>
<td>An infants own thermoregulation is dependent upon skin and vasomotor maturity, improved physical health, fat and glycogen stores, increased body weight and the ability to change position. Also if the infant is unable to maintain own temperature they will require excessive layers of clothing / blankets to allow sufficient observation (St Mary’s Hospital 2006, Fellows 2010). If prolonged TPN is in progress they may have some degree of jaundice.</td>
</tr>
<tr>
<td>Assess infant for suitability:</td>
<td>Age and maturity of infant may have to be taken into account when on TPN. The reason why they are on TPN must also be factored in as in the case of Short bowel syndrome.</td>
</tr>
<tr>
<td>• Desirable weight of &gt; 2 kgs</td>
<td>To maintain the infants’ temperature constant and prevent cold stress (Lyon 2004, St Mary’s Hospital 2006).</td>
</tr>
<tr>
<td>• Five days of consistent weight gain.</td>
<td>To keep parents informed and reduce anxiety and stress caused by hospitalisation (St Mary’s Hospital 2006, Trigg and Mohammed 2010).</td>
</tr>
<tr>
<td>• Clinically well, i.e. no recent history of apnoea, desaturation, bradycardia or convulsions.</td>
<td></td>
</tr>
<tr>
<td>• No evidence of newly acquired or increasing jaundice</td>
<td></td>
</tr>
<tr>
<td>• Ideally not dependent on TPN or other continuous maintenance intravenous fluids</td>
<td></td>
</tr>
<tr>
<td>• Desirable to be fully established on milk feeds (oral / breast) +/- nasogastric / PEG feeds.</td>
<td></td>
</tr>
<tr>
<td>• Infant is able to maintain their own temperature during a nappy change / essential care.</td>
<td></td>
</tr>
<tr>
<td>• Able to maintain own temperature dressed in a single layer of clothing, i.e. babygro, hat and mittens with incubator air temperature set at 30 degree centigrade or less for at least 2 – 3 days prior to transfer.</td>
<td></td>
</tr>
<tr>
<td>Explanation to parents/guardians as clinically indicated.</td>
<td></td>
</tr>
</tbody>
</table>
The cot should be in a draught free area.

The infant should be placed carefully into the cot fully clothed dressed in vest, babygro, cardigan, hat, and gloves with no more than 2 single blankets to maintain temperature as clinically indicated.

The infant should feel comfortably warm to touch all over and not be perspiring.

Baseline observations should be recorded on transfer including both core (tempadot) and skin temperature, heart rate and respiratory rate.

Core and peripheral temperatures should be recorded and documented hourly for 72 hours following transfer from incubator to cot in the PICUs as clinically indicated.

In the ward areas where the infant is more stable / mature record hourly peripheral finger temperatures via touch / feel method for 1-2 days and 2-4 hourly core and toe temperature i.e. "warm to fingers, warm to toes" for 3 days / 72 hours as clinically indicated.

In the ex-PICU graduate infant (prolonged stay) who is stable and has successfully transitioned to a cot, a minimum of 4 hourly monitoring / documenting of core and peripheral temperatures should be undertaken.

If skin temperature drops below 36.5 °C, immediate action should be taken to increase temperature by wrapping up infant and applying more layers of clothing. Record temperature more frequently i.e. at least half hourly as clinically indicated.

Check blood glucose.

If skin temp remains < 36.5 °C despite warming efforts return infant to incubator.

Attempt the whole process again 3 – 4 days later if on repeat assessment the infant is deemed suitable for transfer to a cot i.e. temperature has stabilised and weight gain has continued.

Nests or pillows (under 1 year) should not be used in open cots. If the infant needs to be propped up, put pillow under mattress or tilt the cot if possible.

Avoid bathing infant for approximately . 3-4 days following transfer to the cot until their weight has increased and temperature stabilised in cot and environment.

Ensure that a consistent weight gain is maintained by monitoring weight every 2nd day, post transfer to the cot. Liaise with the dietician as clinically indicated.

To reduce convective heat losses by preventing unnecessary draughts (Trigg and Mohammed 2010).

To maintain a constant the infants temperature constant and prevent cold stress (Lyon 2004). To reduce the incidence of over heating as this is an contributory risk factor in Sudden Infant Death Syndrome (SIDS) (Lyon 2004, Foundation for the Study of Infants Death (FSID) /Department of Health 2009, FSID 2010).

This may indicate overheating.

This will alert staff to changes in temperature and help maintain the infant’s temperature constant and prevent cold stress (Lyon 2004).

Early detection of poor perfusion / cooling of peripheries and timely intervention.

To ensure early and timely intervention for any deterioration in infant’s condition.

To ensure early and timely intervention for any deterioration in infant’s condition.

To reduce convective heat losses and prevent cold stress (St Mary’s Hospital 2006, Fellows 2010).

The infant is at risk of hypoglycaemia when cold stressed.

To maintain neutral thermal environment and prevent cold stress.

This period of time will give the infant time to recover and provides the nurse with valuable time in which to assess the infants overall condition.

As the use of pillows under the age of 1 year increases the incidents of SIDS (Foundation for Sudden Infant Death (FSID) / Department of Health, 2009, FSID 2010). Use of GOI pillows (to prevent or aid plagiocephaly) can be incorporated under strict guidance from Neurodevelopmental Physiotherapists.

Temperature is very unstable during this period. To ensure that energy is not being diverted away from other vital body organs towards maintaining temperature (Lyons 2004, St Mary’s Hospital 2006).

Increased energy expenditure occurs as the infant tries to maintain their body temperature which may affect their weight gain (New et al 2008).
If there are clinical signs of deterioration in the infants general clinical condition i.e.

- increasing apnoea or bradycardia
- sepsis
- milk intolerance
- stop feeding
- convulsions
- metabolic and/or respiratory acidosis

Or any other condition which may require close monitoring and observation, transfer the infant back into a pre-warmed incubator (Section 2 and 3).

Cots are changed weekly for routine cleaning/maintenance. It may also delay the recovery time from other illnesses, etc or may divert energy away from other deprived areas.

To minimise the risk of infection (OLCHC 2006, 2008a).

7.0 References


Coombe Women’s and Infants University Hospital (2009) *Guidelines for Providing a Neutral Thermal Environment for Term and Preterm Infants*. Coombe Woman’s and Infants University Hospital: Dublin.

Coombe Women’s and Infants university Hospital (2010) *Incubator Humidification*. Coombe Women’s and Children’s University Hospital: Dublin.


Our Lady’s Children’s Hospital (2008a) *Infection Control Policy*. OLCHC: Dublin

Queen Mothers Hospital (2009) *Humidity and Care of Humidification systems in the Neonatal Department: Reducing the Risk of Nosocomial Infection*. Queen Mother’s Hospital: Glasgow.


St. Mary’s Hospital (2006) Transferring Infants from Incubators to Open Cots. St Mary’s Hospital: Manchester.

St. Mary’s Hospital (2008) Thermoregulation and the use of Humidity on NNMMU. St Mary’s Hospital: Manchester.


8.0 Appendices

Appendix I

Features of Giraffe

- **Boost Air Curtain**
  This increases the fan speed in order to create a strong blanket of air to serve as a barrier against cool air entry when a porthole or door is opened. It will last for 20 minutes or can be deactivated by the user once the door / porthole is closed returning to “Whisper Quiet” mode (GE Healthcare 2007b, 2010).

- **Phototherapy in the Giraffe:** There is a Giraffe SPOT PT Lite™ (phototherapy) attached to the Giraffe OmniBed which provides intensive phototherapy for infants (Please check the manual for appropriate distance of light from patient) (GE Healthcare 2010).

- **Operating the Giraffe with Two Temperature Probes**
  The OmniBed is equipped with two patient temperature probe jacks for co-bedding of twins, or to monitor a single baby's temperature from two anatomical sites. The OmniBed will only operate in Servo mode from temperature probe 1 (Jack 1). The second temperature probe (Jack 2) may be used in the stable dressed infant who is nursed in air mode to monitor a peripheral temperature (GE Healthcare 2007a).

![Figure 1: Boost Air Curtain on Display.](image1)

![Figure 2: Phototherapy Light.](image2)

Use of Giraffe for Neonates

- **Requiring HFOV**
  Check with engineers in OLCHC re: availability of accessories (Sensormedics HFOV 45-degree fitting or accessory Plexiglas panel) to allow use of HFOV while nursing the infant in Incubator mode.

- **With Gastrochisis silo bag (unrepaired gastrochisis)**
  The Giraffe accessory support arm is available for this purpose. On each of the corners of the translation deck is a hole to mount support arms. The gastrochisis silo may be hung from a support arm mounted through these holes. If the mattress were pulled out, the support arms would move with the baby (GE Healthcare 2007c).
# Appendix II

## Incubator Humidity Record Sheet

**Addressograph**

**Gestational Age**

Age when humidity commenced (if different to GA)

Was this infant in humidity in referring hospital

<table>
<thead>
<tr>
<th>Gestational Age</th>
<th>Starting Level of Humidity</th>
<th>Continued until:</th>
<th>Day 7</th>
<th>Day 11</th>
<th>Day 15</th>
<th>Day 19</th>
<th>Day 23</th>
<th>Day 28</th>
<th>Day 35</th>
<th>Day 42</th>
<th>Day 49</th>
<th>Day 56</th>
<th>Day 63</th>
<th>Day 71</th>
</tr>
</thead>
<tbody>
<tr>
<td>31/40</td>
<td>85%</td>
<td>d/c @ 33 weeks</td>
<td>60%</td>
<td>40%</td>
<td>off</td>
<td></td>
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<td></td>
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</tr>
<tr>
<td>30/40</td>
<td>85%</td>
<td>d/c @ 32 weeks</td>
<td>60%</td>
<td>40%</td>
<td>off</td>
<td></td>
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<tr>
<td>29/40</td>
<td>85%</td>
<td>d/c @ 31 weeks</td>
<td>60%</td>
<td>40%</td>
<td>off</td>
<td></td>
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<td></td>
<td></td>
</tr>
<tr>
<td>28/40</td>
<td>85%</td>
<td>d/c @ 30 weeks</td>
<td>60%</td>
<td>40%</td>
<td>off</td>
<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>27/40</td>
<td>85%</td>
<td>d/c @ 30 weeks</td>
<td>75</td>
<td>65</td>
<td>55</td>
<td>45</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>26/40</td>
<td>85%</td>
<td>d/c @ 32 weeks</td>
<td>80</td>
<td>75</td>
<td>70</td>
<td>65</td>
<td>60</td>
<td>50</td>
<td>40</td>
<td>30</td>
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</tr>
<tr>
<td>25/40</td>
<td>85%</td>
<td>d/c @ 32 weeks</td>
<td>80</td>
<td>75</td>
<td>70</td>
<td>65</td>
<td>60</td>
<td>50</td>
<td>40</td>
<td>30</td>
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</tr>
<tr>
<td>24/40</td>
<td>85%</td>
<td>d/c @ 33 weeks</td>
<td>80</td>
<td>75</td>
<td>70</td>
<td>65</td>
<td>60</td>
<td>55</td>
<td>45</td>
<td>40</td>
<td>30</td>
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<tr>
<td>23/40</td>
<td>85%</td>
<td>d/c @ 33 weeks</td>
<td>85</td>
<td>80</td>
<td>75</td>
<td>70</td>
<td>65</td>
<td>60</td>
<td>55</td>
<td>50</td>
<td>45</td>
<td>40</td>
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</table>

The lowest the humidity can go to on incubator prior to d/c is 30%

<table>
<thead>
<tr>
<th>Date Commenced</th>
<th>Gestational Age</th>
<th>Humidity level</th>
<th>Reduced every 4 days-7 days Date</th>
<th>New level</th>
<th>Discontinuation date</th>
<th>Signature</th>
</tr>
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<tbody>
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</tr>
</tbody>
</table>
Appendix III

Neutral Thermal Environmental Temperatures

Determined by Age and Weight

<table>
<thead>
<tr>
<th>Age</th>
<th>&lt;1200g</th>
<th>1200-2500g</th>
<th>&gt;2500g and &gt;36/40</th>
</tr>
</thead>
<tbody>
<tr>
<td>0-24hrs</td>
<td>34.0-35.4</td>
<td>31.5-34.0</td>
<td>31.0-33.8</td>
</tr>
<tr>
<td>24-48hrs</td>
<td>34.0-35.0</td>
<td>31.5-34.0</td>
<td>30.5-33.5</td>
</tr>
<tr>
<td>48-96hrs</td>
<td>34.0-35.0</td>
<td>31.0-34.0</td>
<td>29.8-33.2</td>
</tr>
<tr>
<td>4-14 days</td>
<td>32.6-34.0</td>
<td>30.0-33.0</td>
<td>29.0-32.6</td>
</tr>
<tr>
<td>2-6 weeks</td>
<td>30.6-34.0</td>
<td>29.0-32.0</td>
<td>29.0-33.0</td>
</tr>
</tbody>
</table>

(Hazinski 1999, Blumer 1990)